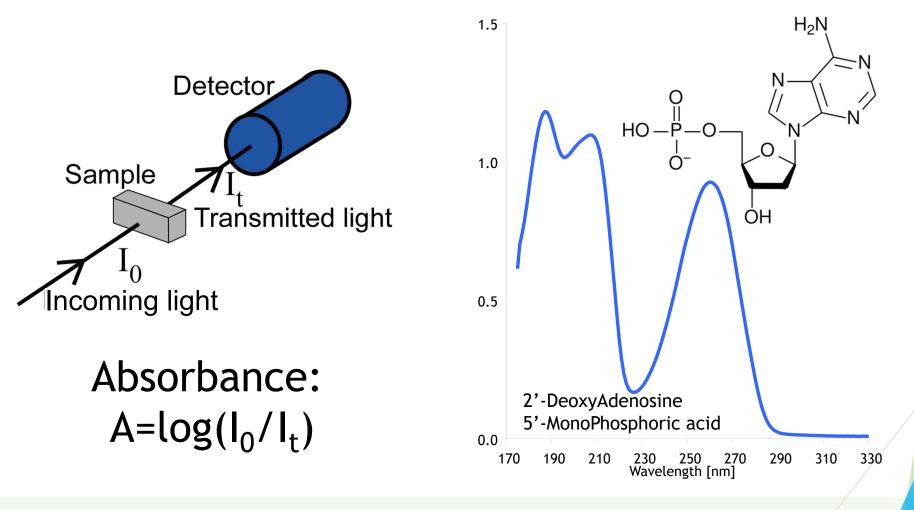


ESC1: Circular Dichroism: best practice and data analysis

Lecture 2: Instrumentation

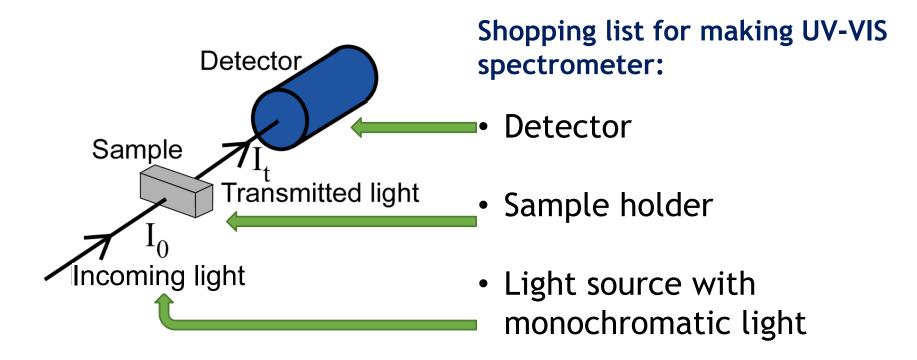


Absorption Spectroscopy





UV-VIS Instrumentation



A monochromatic source means a single wavelength of light

➡ To make such a source we need a *lamp* and *optics*



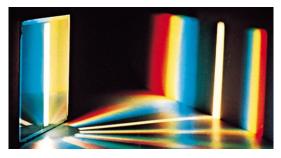
UV-VIS Instrumentation

<u>Mirrors</u>

Plane mirrors

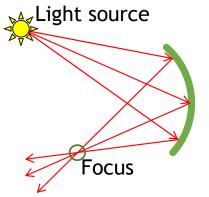


Dispersing elements





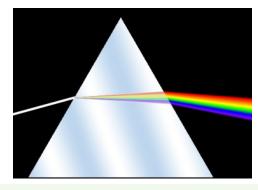
Curved mirrors



Prism

Diffraction

grating

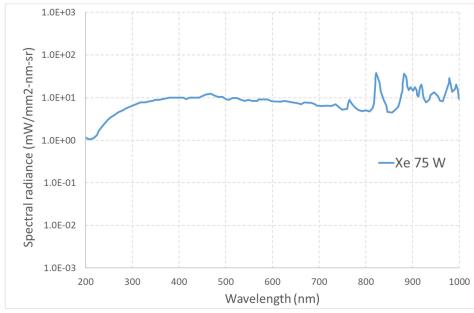






UV-VIS Instrumentation: Light source

Xenon arc lamp







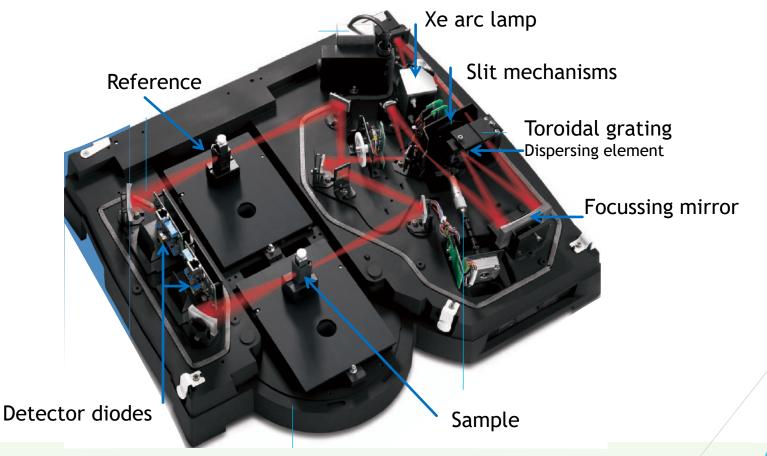


Quite intense UV lamp

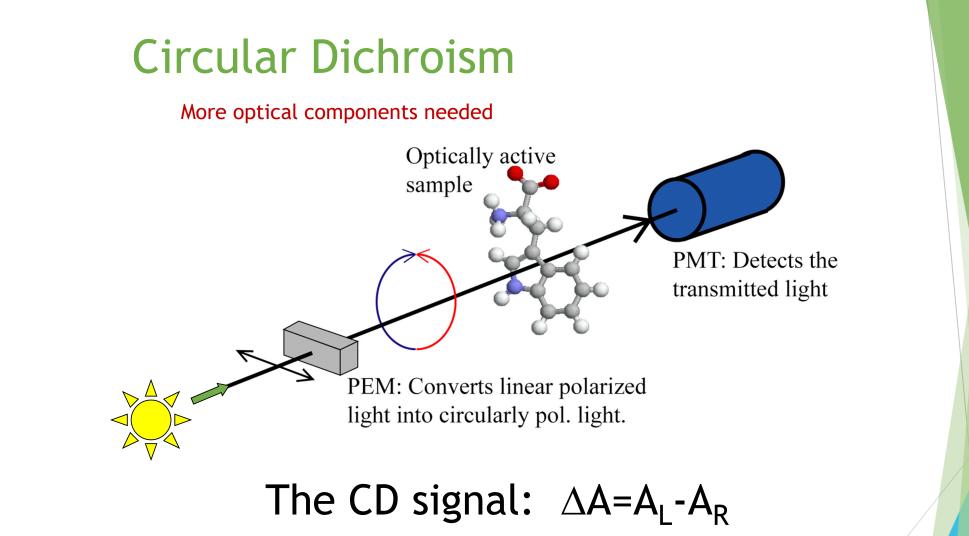


UV-VIS Instrumentation

Conventional UV-VIS instruments

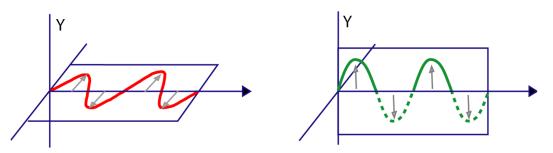




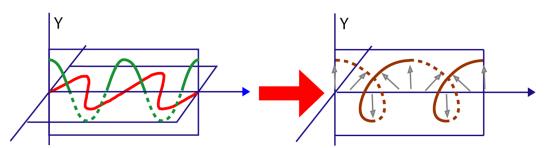




Polarized light



Horizontally pol. Vertically pol.



Sum of two plane pol. Circularly pol.

How do we change the polarisation of light?

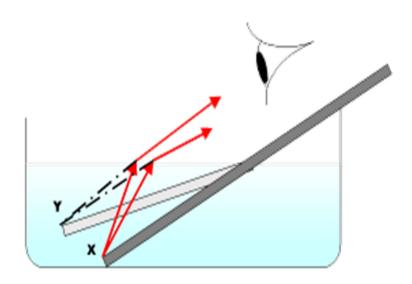




Refraction

Refraction: Light is refracted when travelling from one medium to the next



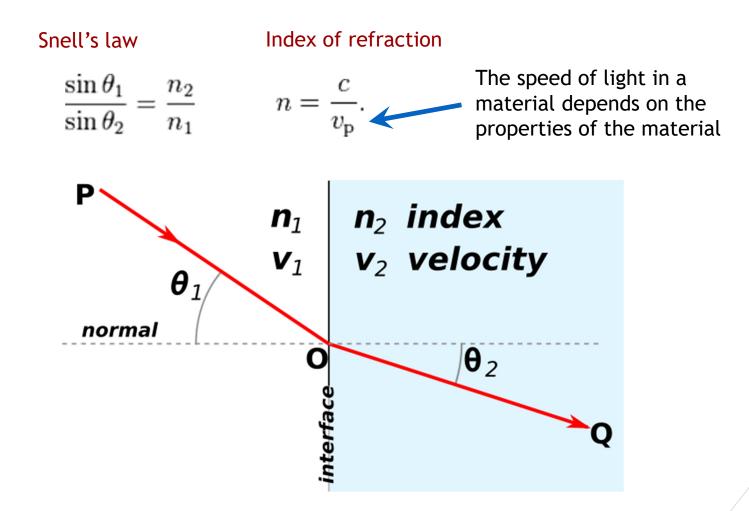


This is (partly) why it's difficult to catch a fish in water

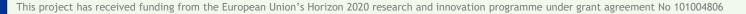




Refraction





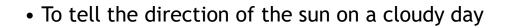


Birefringence

Birefringence, or double refraction



Icelandic Spar / Calcite / CaCO₃ Used by the Vikings for navigation (sunstone)









What is Birefringence?

Indexes of refraction depend on polarization

The birefringent effect (using calcite) was first described by scientist Rasmus Bartholin in 1669

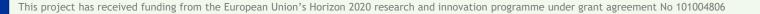
If a crystal has two different Indexes of refraction: $n_{\rm e}$ and $n_{\rm o}$

Birefringence $\Delta n = n_e - n_o$



Calcite: $\Delta n(590 \text{ nm}) = 1.486 - 1.64 = -0.154$





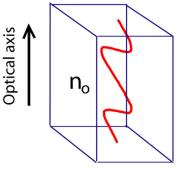
What is Birefringence

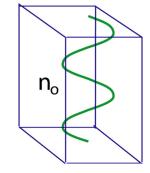
What is meant by two indexes of refraction?

The material (crystal) has a direction: the optical axis

Light travelling along the optical axis:

n independent of polarization

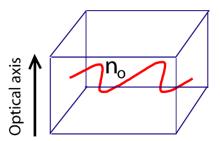


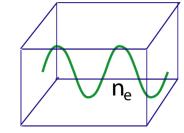


Light travelling perpendicular to the optical axis: n dependents on polarization

Pol. Loptical axis: n_o

Pol. || optical axis: n_e



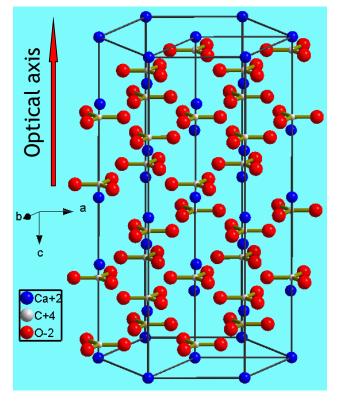






Origin of Birefringence

Crystal structure of Calcite



Calcite is an uniaxial crystal

' **n**e Birefringence $\Delta n = n_{\rm P} - n_{\rm o}$

o MOSE

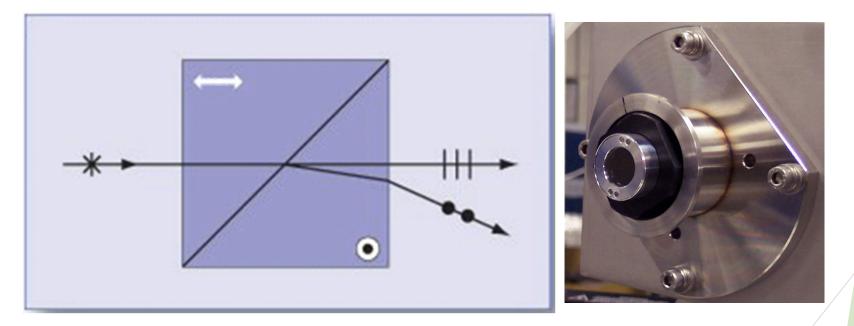


The Rochon Polarizer

"MgF2 is <u>slightly</u> birefringent..."

- Info on MgF₂ from Crystran Ltd., UK.

MgF₂: $\Delta n(546nm) = 1.390 - 1.379 = 0.011$ (<< $\Delta n_{Calcite}$)







Retarders/Wave plates

Optical axis

- The speed of the light, and thus the wavelength, is changed inside the crystal
- The horz. pol. light is slightly faster than the vert. pol. light.
- The initially 45 deg. pol. light 'flips' the polarization by 90 deg.

Retardation R = $(n_e - n_o) d / \lambda$

¹/₂ waveplate



Retarders/Wave plates

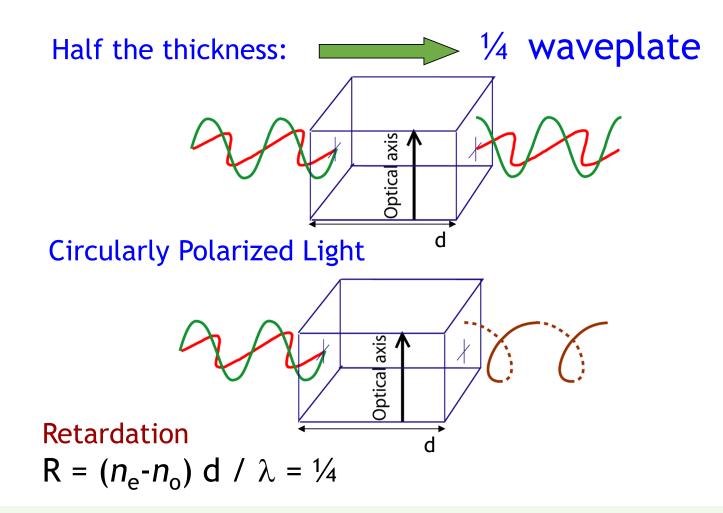
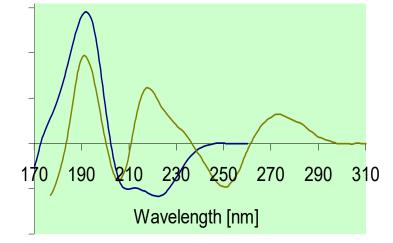




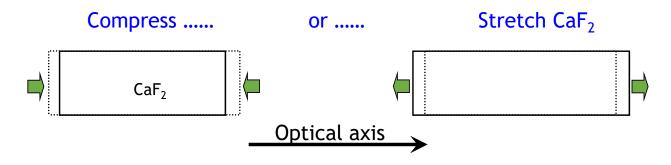


Photo Elastic Modulators (PEM)

- We are scanning the wavelength
- We can't change d as we scan



To keep R = 1/4 (circularly pol) change n_e - n_o by:







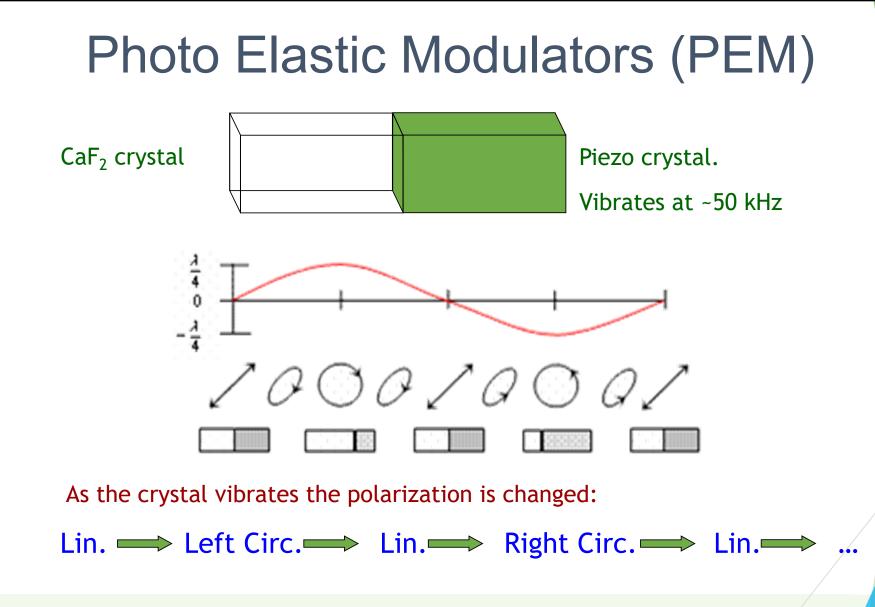
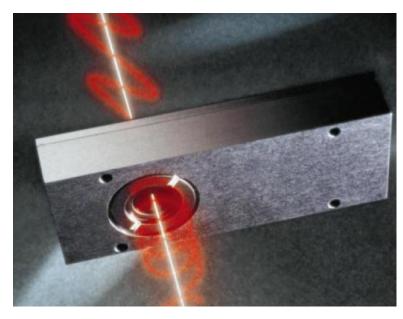




Photo Elastic Modulators (PEM)



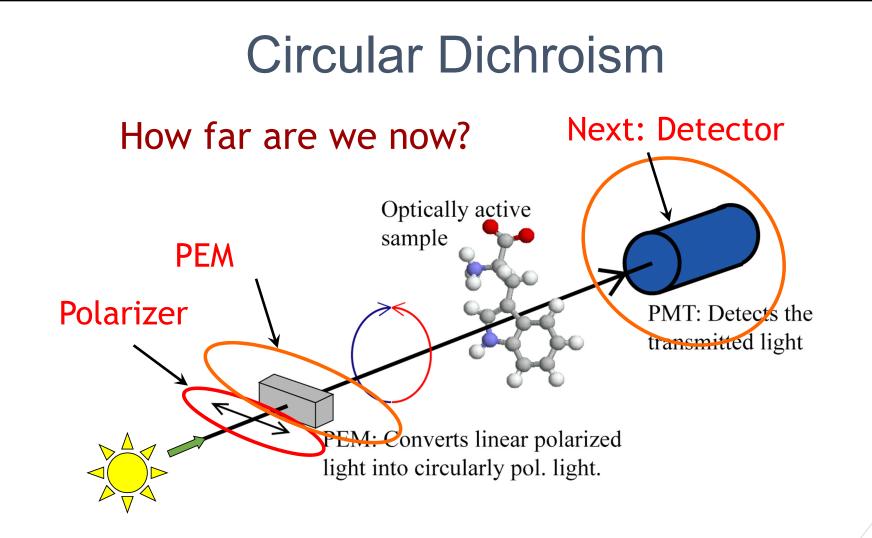
Linear pol. is converted to Circ. pol.



CaF₂ crystal

Piezo crystal. Vibrates at ~50 kHz



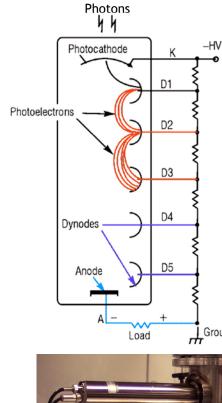


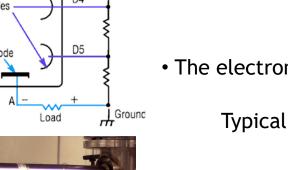




CD/UV-VIS Instrumentation

Photomultiplier Tube (PMT) Detector

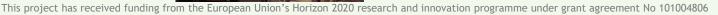






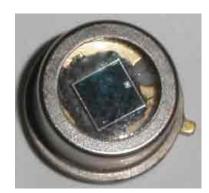
- A photon is converted to an electron on the cathode: Q_{eff} (quantum efficiency)
- The cathode can be optimized to certain wavelength ranges UV, VIS or even Solar Blind
- A high voltage (HV) drop is applied along the dynodes
- The photon electron is amplified to many electrons: Gain(HV) (strongly depends on HV) Gain(HV) $\approx b \cdot (HV)^a$
- The electrons are collected on the Anode: Either pulses or a current is detected Typically: Gain(HV) $\approx 10^3 - 10^7$ and Q_{eff}<1



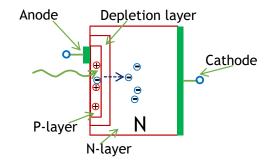


CD/UV-VIS Instrumentation

Photodiode



Cathode Anode

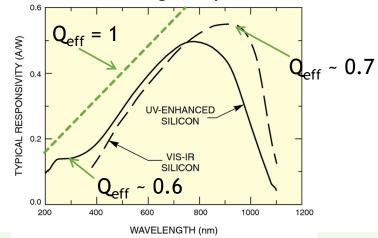


- The diode converts photons to a current
- Q_{eff} can be as high as 80%
- Spectral range depends on material E.g. Si 190 - 1100 nm, Ge 400 - 1700 nm

Advantage:

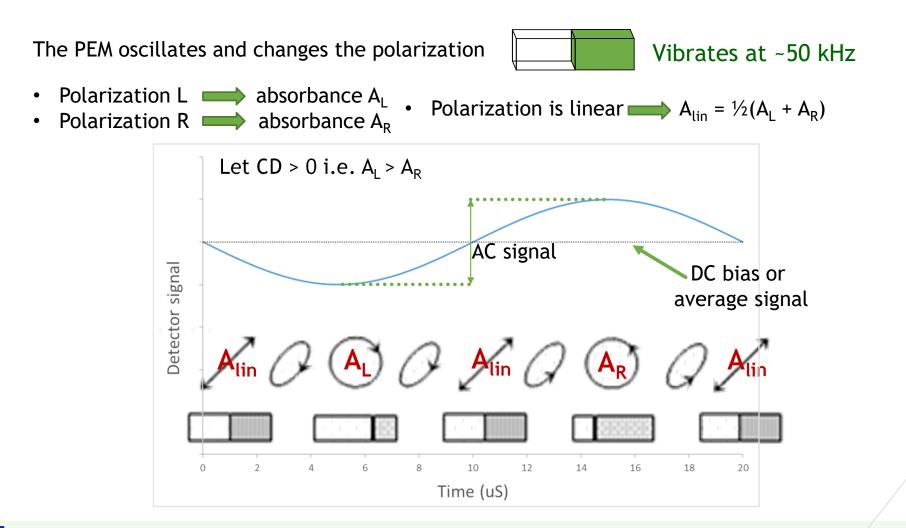
Very rugged, high Q_{eff} (in the VIS/IR) <u>Disadvantage:</u>

No gain , although avalanche diodes can have a gain up to 1000





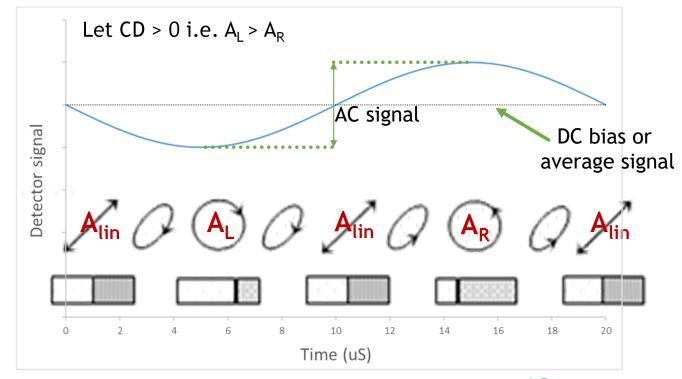
The signal from the detector







The signal from the detector



The CD instrument measures the AC (and DC)

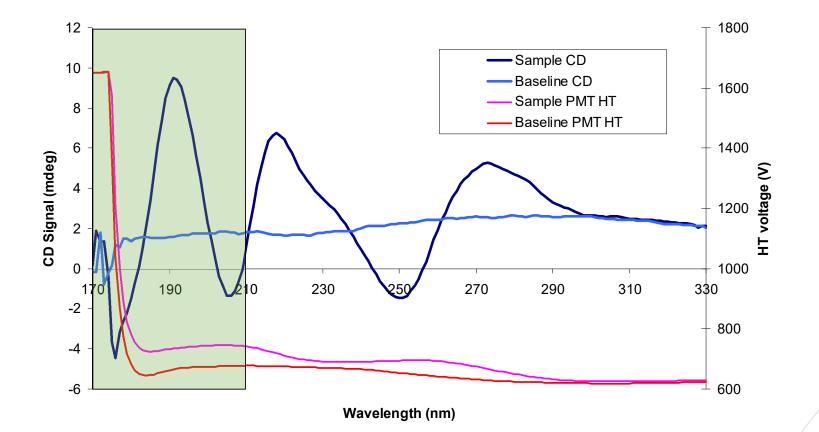
 $CD = constant \times \frac{AC}{DC}$

In most instruments the DC is kept constant by changing the detector high voltage (HT)

$$CD = const \times AC$$

What does the HT tell us?



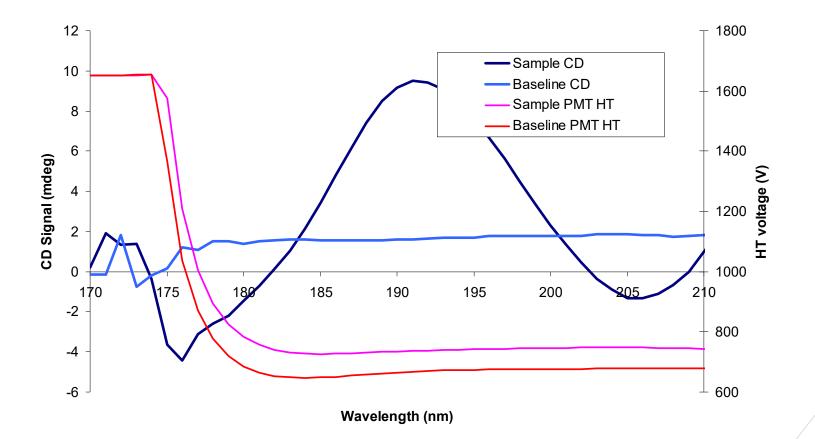




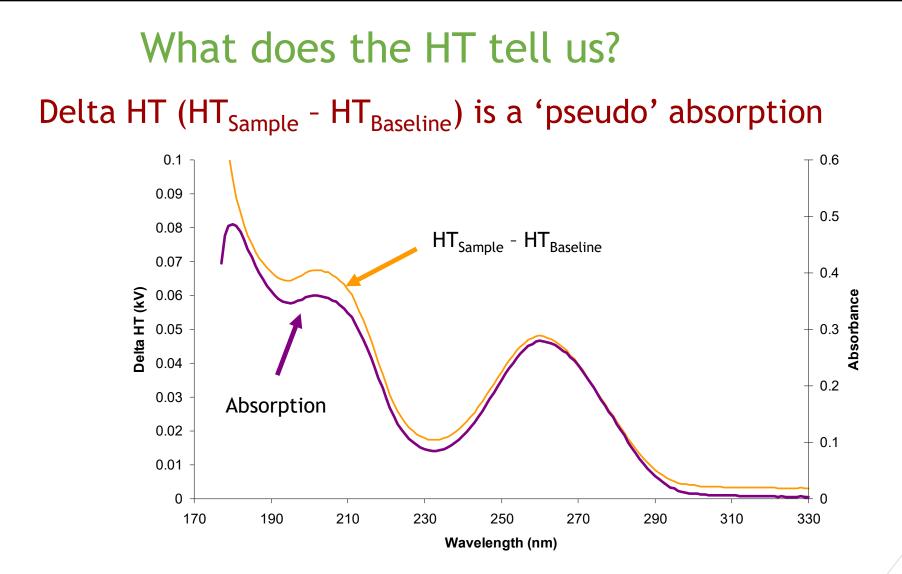


What does the HT tell us?

High HT 🛑 Low photon flux reaching detector 🛑 High Absorbance









What does the HT tell us?

It is possible to use HT_{Sample} and $HT_{Baseline}$ to calculate the absorbance

- Record the HT_{Sample} (λ) and $HT_{Baseline}$ (λ) together with the CD signal
- The average signal from the detector is constant (DC bias)
- Assume that for sample and baseline these are the same:
 - Lamp output
 - Optics transmission
- Assume the gain of the detector vs. HT is $Gain(HT) = b \times HT^{a}$

 $Detector signal_{sample} = Detector signal_{baseline}$

$$10^{-A_{sample}(\lambda)} \times Gain(HT_{sample}(\lambda)) = 10^{-A_{baseline}(\lambda)} \times Gain(HT_{baseline}(\lambda))$$

<u>Use Gain(HT) = $b \times HT^{a}$ </u>: $10^{A_{sample}(\lambda) - A_{baseline}(\lambda)} = \frac{b \times (HT_{sample}(\lambda))^{a}}{b \times (HT_{baseline}(\lambda))^{a}}$

$$A(\lambda) = a \times \log\left(\frac{HT_{sample}(\lambda)}{HT_{baseline}(\lambda)}\right)$$

Only depends on a **single constant** *a* ! Calibrate this using samples with know concentrations

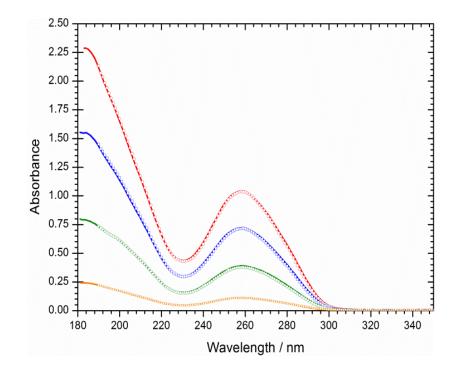


What does the HT tell us?

Highly accurate **simultaneous** CD and Absorption measurements

$$A(\lambda) = a \times \log\left(\frac{HT_{sample}(\lambda)}{HT_{baseline}(\lambda)}\right)$$

Full range of absorptions 0-2 calibrated with a **single** detector parameter

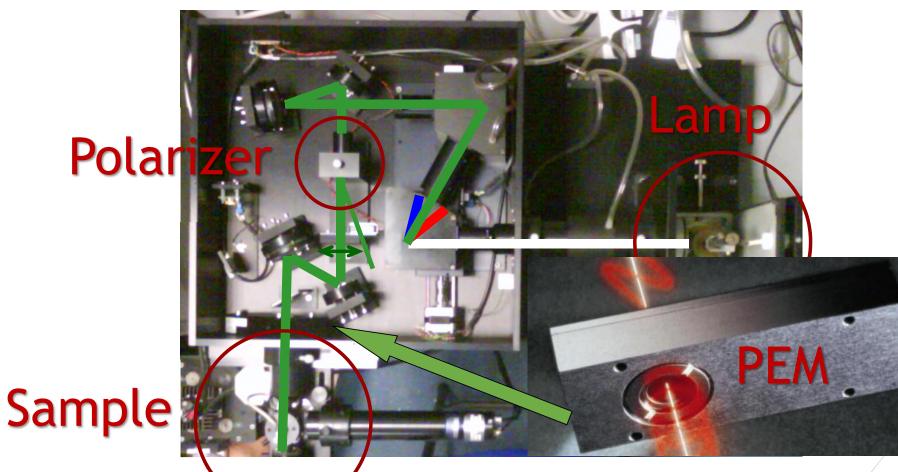






Instrumentation

Conventional CD instrument (lamp based)

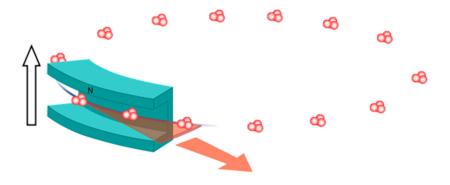


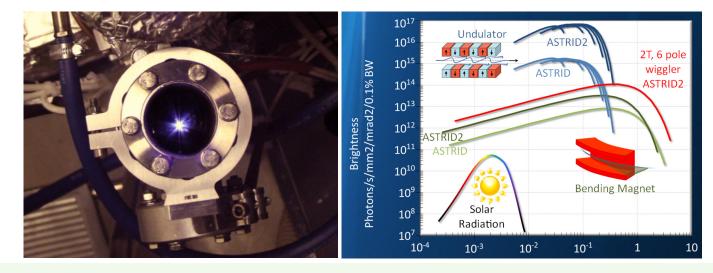




Other options for light sources: Synchrotron Radiation (SR)

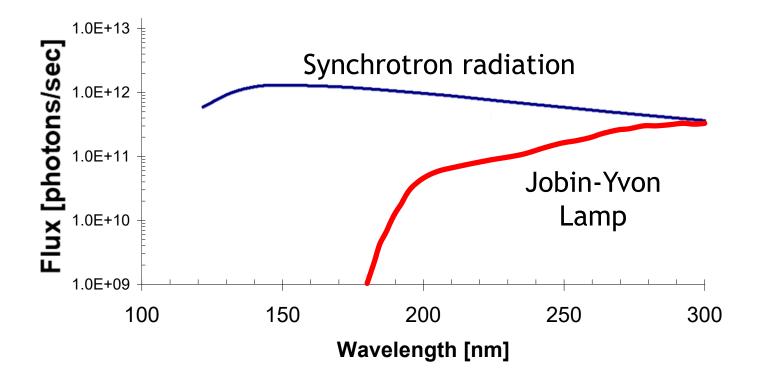
- UV light is well suited to examine molecules like proteins and DNA.
- Synchrotron radiation (SR) is emitted when charged particles are accelerated: We use electrons at relativistic speeds.
- The light is VERY intense.



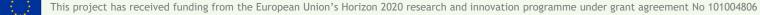




Other options for light sources: SR vs lamps

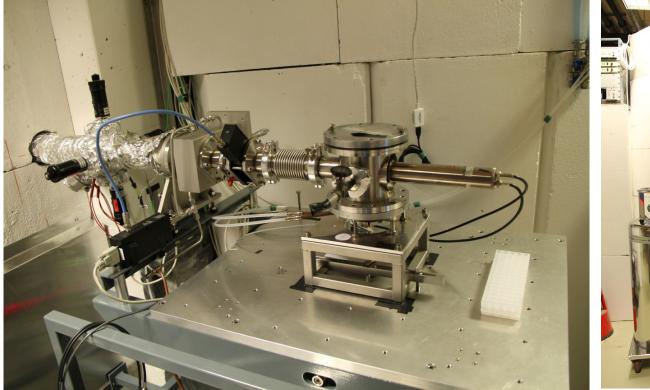






AU-CD beam line on ASTRID2

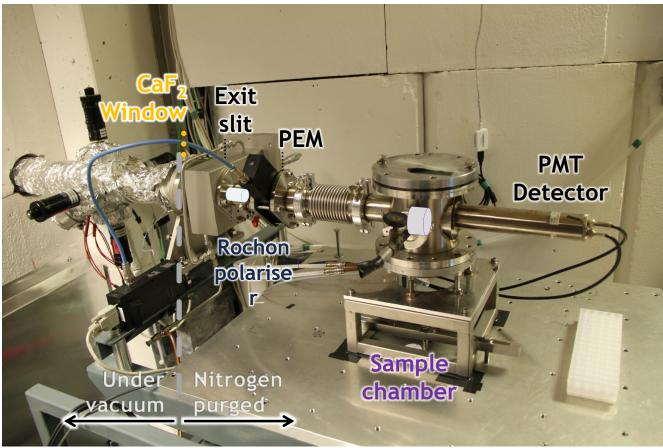
Small and compact set-up







AU-CD beam line on ASTRID2









Sample holders - solvents Cells made from Quartz suprasil with a cut-off of ~160 nm







Sample holders - solvents Cells made from Quartz suprasil with a cut-off of ~160 nm

We prefer to use round cells as they often have less stress and thus birefringence





- Closed cells
 - Pathlengths 0.1 mm to several cm
 - Temperature melts

- Open cells
 - Shorter pathlengths (~0.01 mm or lower!)
 - CaF₂ cells, lower wavelength cutoff



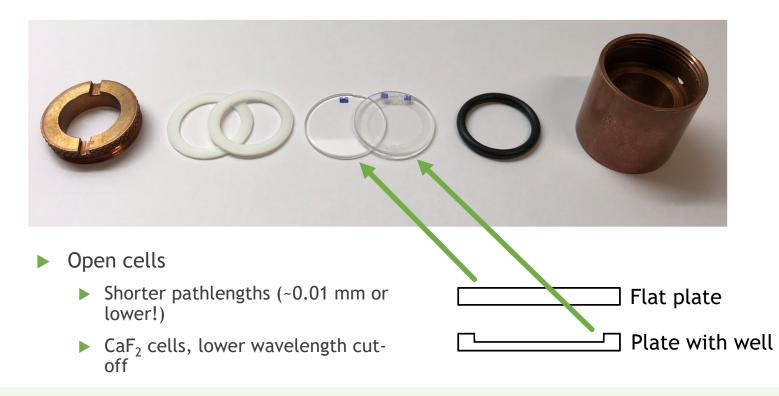




Sample holders - solvents

Cells made from Quartz suprasil with a cut-off of ~160 nm

We prefer to use round cells as they often have less stress and thus birefringence





A note on cell cleaning

Some cells are easier to clean than others

QS 10.00 mm

Large open cuvette: Not so difficult to clean <u>Use:</u> Water



Open cells: Easy to clean <u>Use:</u> Water and tissue



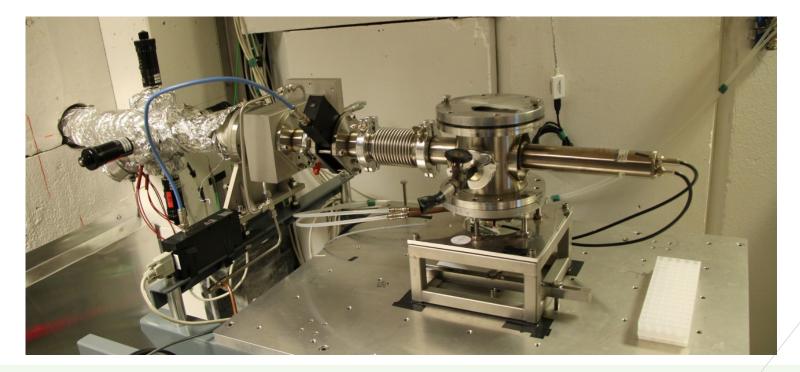
Closed cells: Difficult to clean <u>Use:</u> Hellmanex 2% or 10% solution and heat

Much more about this in the first Hands-on session



Measurement options...

Temperature scans 5 to 90C - fully automated and integrated into the scanning programme using a macro file.

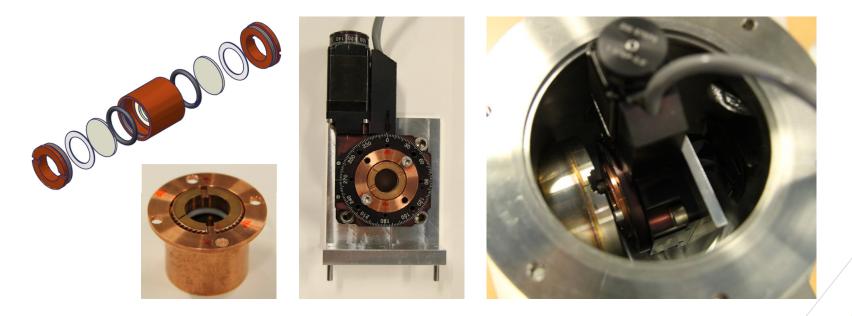






Measurement options...

- Temperature scans 5 to 90C fully automated and integrated into the scanning programme using a macro file.
- Rotational stage

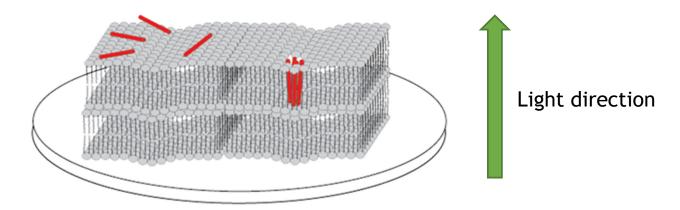






Why a rotation sample holder?

- Often used when studying films (solids) of samples
- ► A good example is insertion of peptides into lipid bilayers



Lipid bilayers (mixed with peptides) can be made on e.g. a quartz plate by drying a solution onto the plate.

Are the peptides inserted into the membrane or are they surface bond?

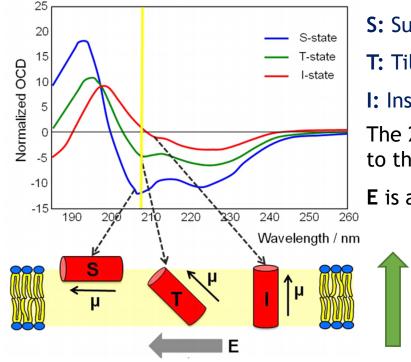
This is a very different sample than a solution where the peptides are found in all directions with respect to the light



Why a rotation sample holder?

Oriented Circular Dichroism (OCD)

The CD spectrum of an helical peptide in the lipid bilayer depends on the orientation of the peptide.



S: Surface bond peptide

T: Tilted peptide

I: Inserted peptide

The 208 nm transition (μ) is only active when parallel to the light electric field, E ($\mu \parallel E$)

E is always perpendicular to the direction of the light

Light direction

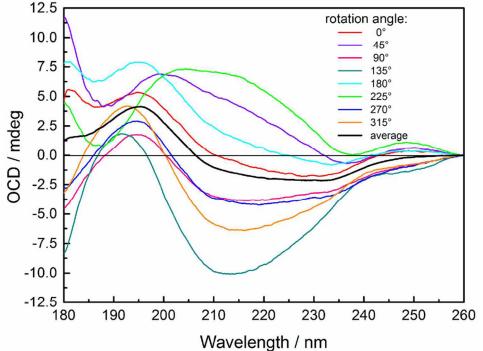
J. Bürck et al. Acc. Chem. Res. 2016, 49, 184–192

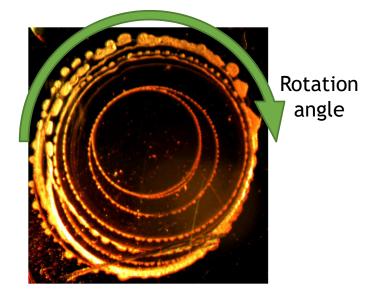


Why a rotation sample holder? Oriented Circular Dichroism (OCD)

Such a (solid) sample is not necessarily very uniform, and may be a bit isotropic

Shows up as changes in the CD spectrum with rotation angle

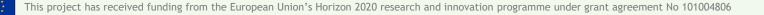




A not particularly uniform sample!

The CD signal depends on sample angle, but averages to the correct spectrum!



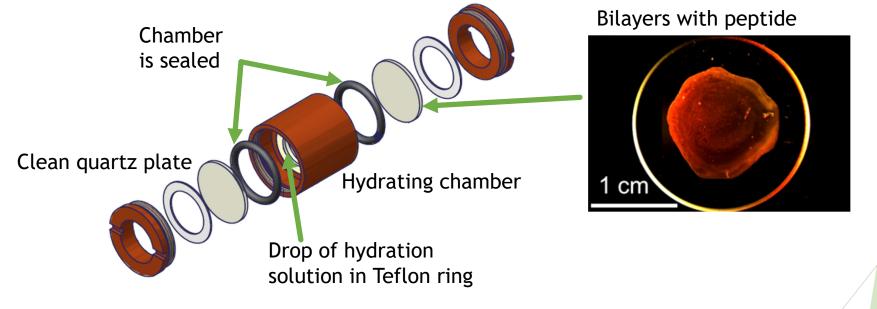


Why a rotation sample holder?

Oriented Circular Dichroism (OCD)

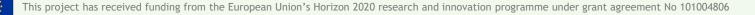
One important aspect of peptides in a bilayer is that they need to be kept humid and might also need to be hydrated after preparation for the peptides to insert into the membrane.

Special sample holder



The humidity can be controlled with different salt solutions: K_2SO_4 R.H. ~98 % MgCl₂ R.H. ~ 33 %

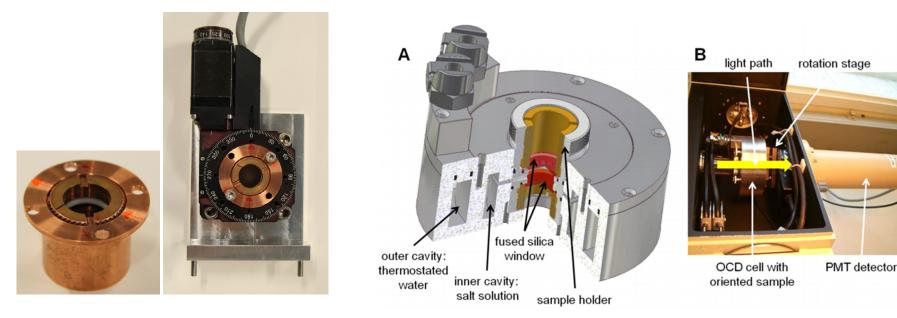




Why a rotation sample holder? Oriented Circular Dichroism (OCD)

Our setup for rotation

KIT (DE) setup for a JASCO CD instrument

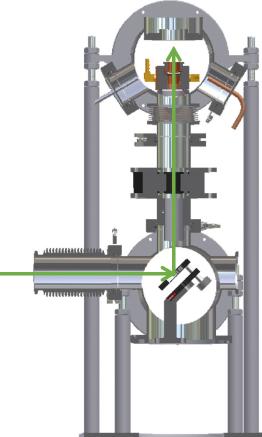


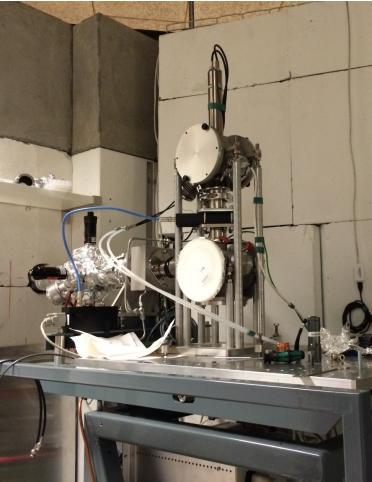
The hydrating sample holder is mounted in the rotational stage, and CD spectra are (automatically) acquired at many different angles.



Measurement options... Periscope

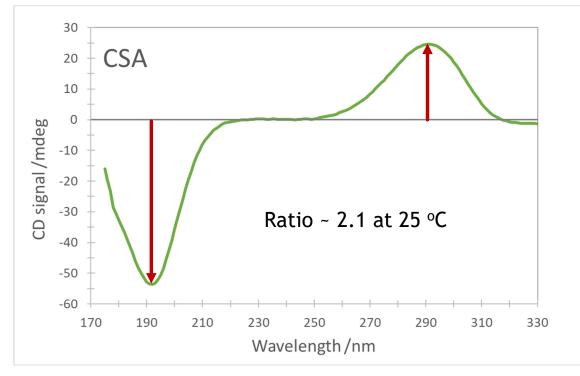
The periscope allows samples to be measured in a horizontal position







As with any other instrument, it is vital that you keep your instrument well calibrated Check the calibration at the start of each day using (1S)-(+)-10-camphorsulfonic acid (CSA)



 $\Delta \epsilon_{290.5nm} = 2.36 \text{ M}^{-1} \text{ cm}^{-1}$ MW = 232.30 g/mol Use ~7 mg/ml in 0.1 mm cell

CSA concentration:

- CSA is hydroscopic
- Measure A_{285nm} in 1 cm cuvette
- Use $\epsilon_{285} = 34.6 \text{ M}^{-1} \text{ cm}^{-1}$
- Rule of thumb:

A₂₈₅(1cm) = 1 for 6.71 mg/ml

Ratio must be above 2. Ratio is very temperature dependent: always use 25°C.



With time any spectrophotometer may develop stray light

- Stray light may pass through the sample
- If the absorption is high, the stray light hitting the detector may be significant
- The HT will become too low when keeping DC bias constant
 - Leads to CD features being measured too low!

The absorbance at the 192 nm CSA peak is relatively high

> Stray light shows up as the 192 / 290 ratio becoming low

Daily measurements of the CSA peak values gives a track record of the health of the instrument:

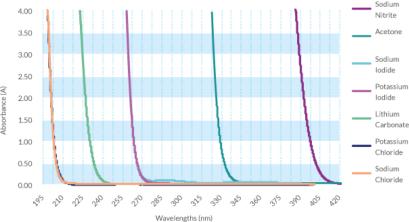
- The 192 / 290 ratio gives information about stay light changes over time
- The value of the 290 nm CSA CD signal gives information about the absolute calibration



With time any spectrophotometer may develop stray light

E.g. Starna has reference salts for measuring stray light





Stray Light Cut-off Spectra

Or you can make your own 12 g/l KCl solution:

> Absorbance > 2.0 measured at 198 nm against a water reference





Check the wavelength calibration

- Daily by checking the position of the two CSA peaks (a rough estimate)
- Every few months using Holmium Oxide (HoOx)
 - 40 g/L solution of holmium oxide in 10% (v/v) perchloric acid
 - Store in sealed 10 mm cuvette

