

Protein Unfolding Data Analysis

DSC & DSF

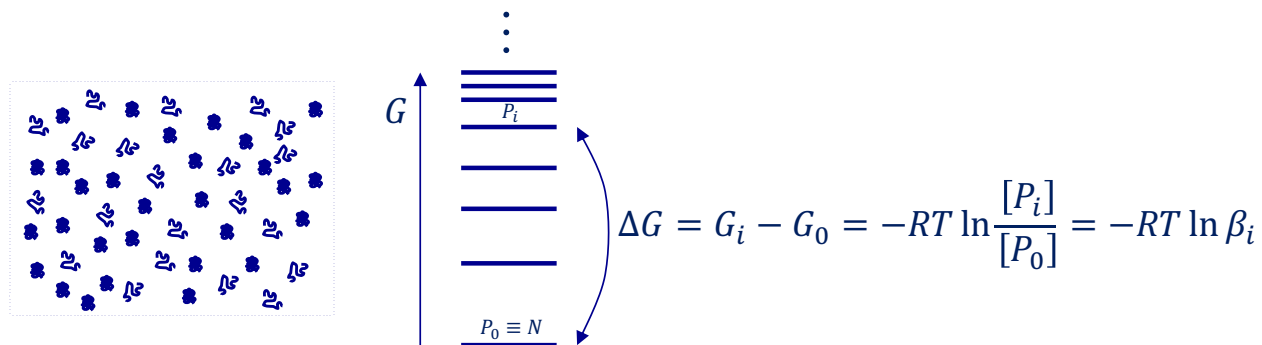
Adrian Velazquez-Campoy



This project has received funding from the European Union's Horizon 2020 research and innovation programme under grant agreement No 101004806

Stability

ΔG Gibbs energy of stabilization or unfolding
Difference in Gibbs energy between the native state
and the unfolded state



► Standard approach based on partition function Q

$$P_0 \leftrightarrow P_1 \leftrightarrow P_2 \leftrightarrow \cdots \leftrightarrow P_i \leftrightarrow \cdots \leftrightarrow P_n$$

$$[P_i] = \beta_i [P_0]$$
$$[P_i] = \left(\prod_{r=1}^i K_r \right) [P_0]$$

$$Q = \sum_{i=0}^n \frac{[P_i]}{[P_0]} = \sum_{i=0}^n \beta_i$$

Wyman & Gill. "Binding and Linkage", University Science Books, 1990



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► Standard approach based on partition function Q

$$Q = \sum_{i=0}^n \frac{[P_i]}{[P_0]} = \sum_{i=0}^n \beta_i$$

$$\chi_i = \frac{[P_i]}{[P]_T} = \frac{\beta_i}{Q}$$

$$\beta_i = \exp(-\Delta G_i/RT)$$

$$\Delta G_i(T) = \Delta H_i(T_{m,i}) \left(1 - \frac{T}{T_{m,i}}\right) + \Delta C_{P,i} \left(T - T_{m,i} - T \ln \frac{T}{T_{m,i}}\right)$$

$$\Delta G_i = \Delta G_i(T, pH, \mu, [D], [L], a_w, \dots)$$



► Standard approach based on partition function Q

$$Q = \sum_{i=0}^n \frac{[P_i]}{[P_0]} = \sum_{i=0}^n \beta_i$$

$$\chi_i = \frac{[P_i]}{[P]_T} = \frac{\beta_i}{Q}$$

$$\beta_i = \exp(-\Delta G_i/RT)$$

$$\langle S \rangle = \sum_{i=0}^n \chi_i S_i = \frac{\sum_{i=0}^n \beta_i S_i}{\sum_{i=0}^n \beta_i}$$

**Average of any system property
(measurement)**

$$NLSF \rightarrow \{T_{mi}, \Delta H_i, \Delta C_{Pi}; i = 1, \dots, n\}$$



► Standard approach based on partition function Q

$$Q = \sum_{i=0}^n \frac{[P_i]}{[P_0]} = \sum_{i=0}^n \beta_i \quad \chi_i = \frac{[P_i]}{[P]_T} = \frac{\beta_i}{Q}$$

$$\langle \Delta H \rangle = RT^2 \frac{\partial \ln Q}{\partial T} = \frac{\sum_{i=0}^n \beta_i \Delta H_i}{\sum_{i=0}^n \beta_i} \quad \beta_i = \exp(-\Delta G_i/RT)$$

$$\langle \Delta C_P \rangle = \left(\frac{\partial \langle \Delta H \rangle}{\partial T} \right)_P = \langle \Delta C_P \rangle_{int} + \frac{\langle \Delta H^2 \rangle - \langle \Delta H \rangle^2}{RT^2} \cong \frac{\langle \Delta H \rangle (T + \Delta T) - \langle \Delta H \rangle (T)}{\Delta T}$$

$$NLSF \rightarrow \{T_{mi}, \Delta H_i, \Delta C_{Pi}; i = 1, \dots, n\}$$



Single-transition unfolding



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$$N \leftrightarrow U$$

i	G	ΔG	$\exp(-\Delta G/RT)$
fold	G_N	0	1
unfold	G_U	ΔG	K

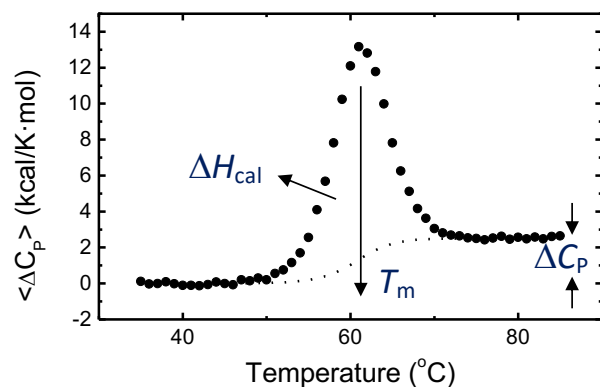
$$Q = 1 + \beta = 1 + K$$

$$\chi_N = \frac{1}{1 + K}$$

$$\chi_U = \frac{K}{1 + K}$$



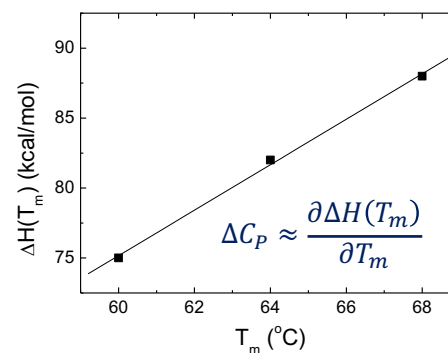
► Single transition unfolding: model-free analysis

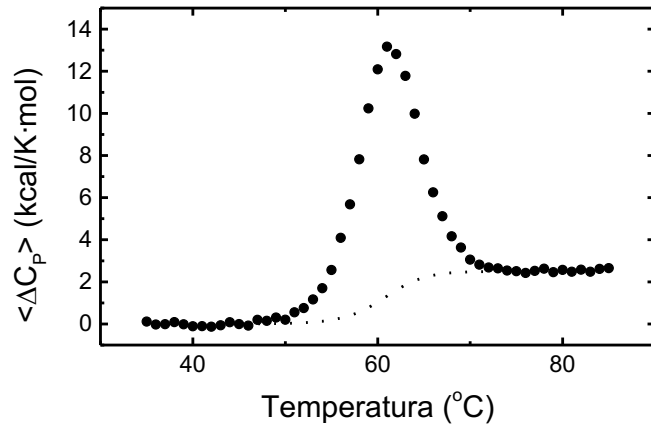


- T_m and ΔH : well determined
- ΔC_p : poorly determined

$$\Delta H(T_m) = \int_{T_0}^{T_1} \langle \Delta C_p \rangle dT$$

$$\Delta S(T_m) = \int_{T_0}^{T_1} \frac{\langle \Delta C_p \rangle}{T} dT$$





$$\langle \Delta H \rangle = \chi_N \Delta H_N + \chi_U \Delta H_U = \chi_U \Delta H$$

$$\Delta H(T) = \Delta H(T_m) + \Delta C_P (T - T_m)$$

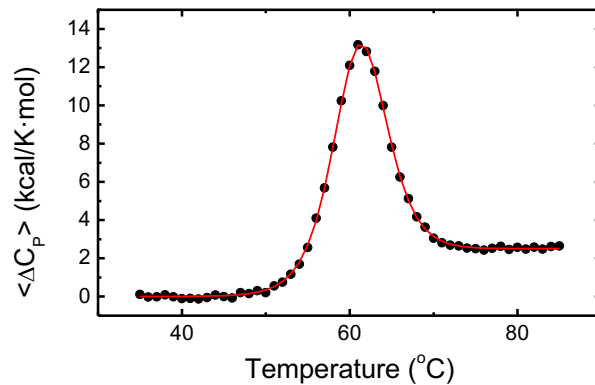
$$\Delta G(T) = \Delta H(T_m) \left(1 - \frac{T}{T_m}\right) + \Delta C_P \left(T - T_m - T \ln \frac{T}{T_m}\right) \quad K = e^{-(\Delta G(T)/RT)}$$

$$\chi_N = \frac{[P_0]}{[P]_T} = \frac{1}{1 + K} \quad \chi_U = \frac{[P_1]}{[P]_T} = \frac{K}{1 + K}$$

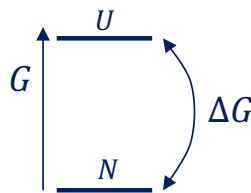
$$\langle \Delta C_P \rangle = \frac{K}{(1 + K)^2} \frac{\Delta H^2}{RT^2} + \frac{K}{1 + K} \Delta C_P \cong \frac{\langle \Delta H \rangle (T + \Delta T) - \langle \Delta H \rangle (T)}{\Delta T}$$



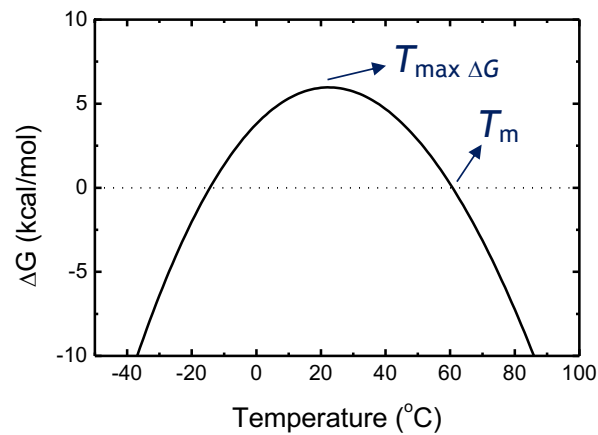
► Single transition unfolding: NLSF analysis



$$\begin{aligned} T_m &= 60.75 \pm 0.03 \text{ }^\circ\text{C} \\ \Delta H(T_m) &= 102 \pm 0.3 \text{ kcal/mol} \\ \Delta C_p &= 2.51 \pm 0.02 \text{ kcal/K}\cdot\text{mol} \end{aligned}$$



Stability profile $\Delta G(T)$



Two-transition unfolding



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i	ΔG_i	$\exp(-\Delta G_i/RT)$
	0	1
	ΔG_1	K_1
	ΔG_2	K_2
	$\Delta G_1 + \Delta G_2$	$K_1 K_2$

$$Q = 1 + \beta_1 + \beta_2 + \beta_3 = 1 + K_1 + K_2 + K_1 K_2$$

$$\chi_N = \frac{1}{1 + K_1 + K_2 + K_1 K_2} \quad \chi_{I_2} = \frac{K_2}{1 + K_1 + K_2 + K_1 K_2}$$

$$\chi_{I_1} = \frac{K_1}{1 + K_1 + K_2 + K_1 K_2} \quad \chi_U = \frac{K_1 K_2}{1 + K_1 + K_2 + K_1 K_2}$$



i	ΔG_i	$\exp(-\Delta G_i/RT)$
	0	1
	ΔG_1	K_1
	$\Delta G_1 + \Delta G_2$	$K_1 K_2$

$$Q = 1 + \beta_1 + \beta_2 = 1 + K_1 + K_1 K_2$$

$$\chi_N = \frac{1}{1 + K_1 + K_1 K_2}$$

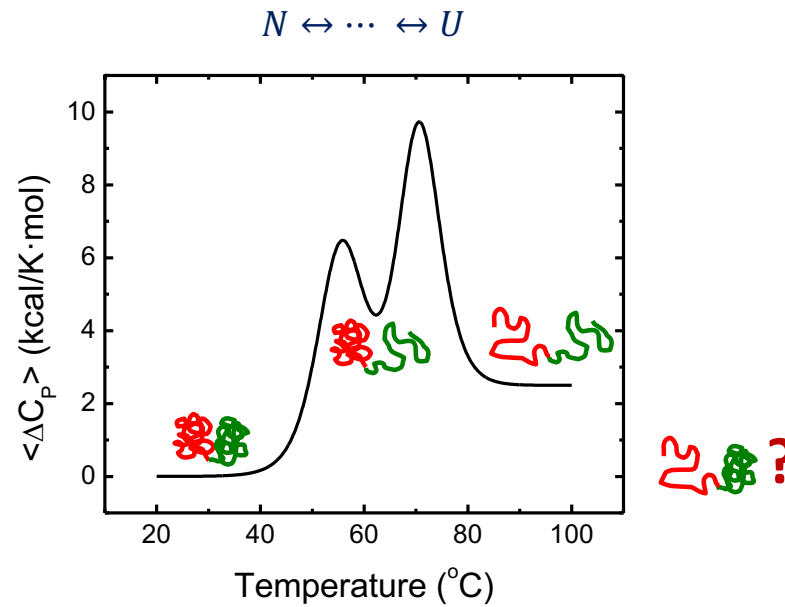
$$\chi_{I_2} \approx 0$$

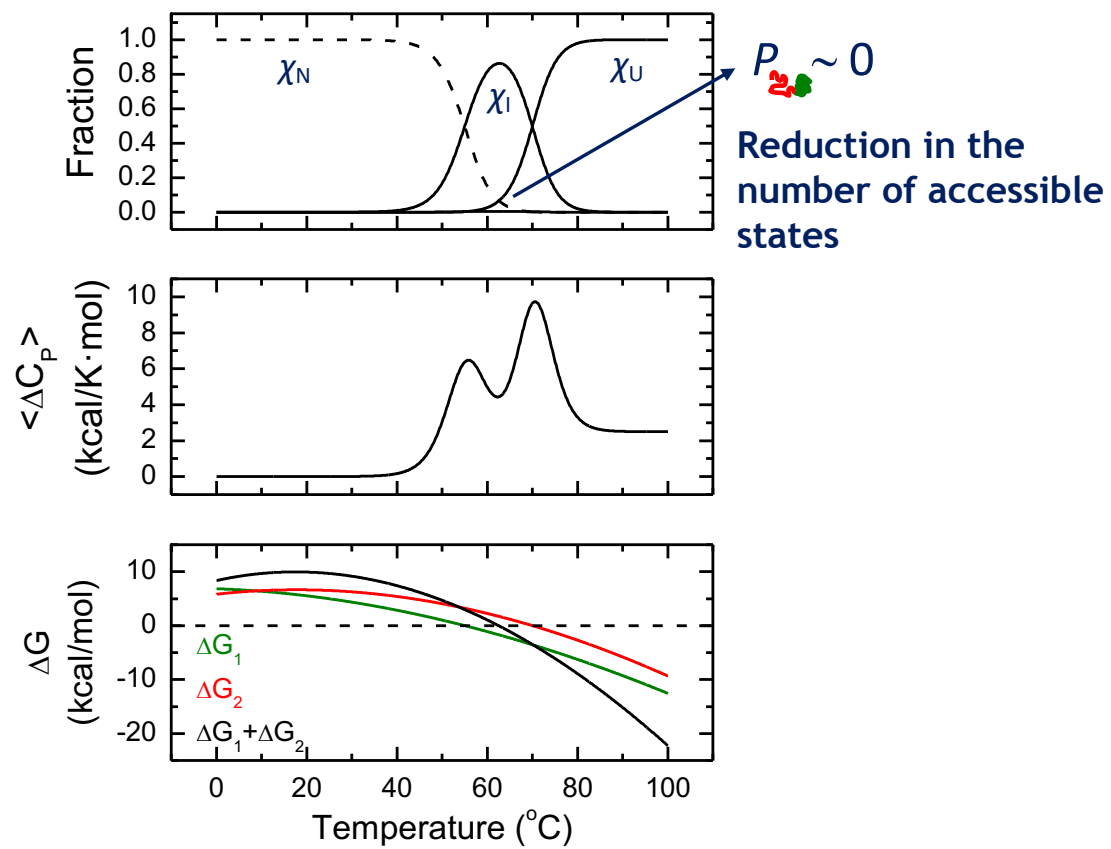
$$\chi_{I_1} = \frac{K_1}{1 + K_1 + K_1 K_2}$$

$$\chi_U = \frac{K_1 K_2}{1 + K_1 + K_1 K_2}$$



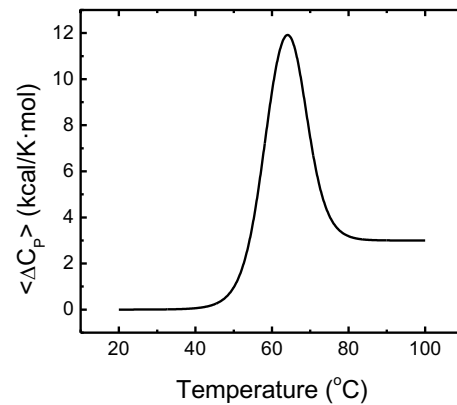
► Two-transition unfolding (three-state unfolding?)





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► How many transitions?



Two-state test (single transition test):

- Overlapping of unfolding curves obtained with different techniques (spectroscopy)
- van't Hoff-calorimetric enthalpies ratio (calorimetry) $\frac{\Delta H_{vH}}{\Delta H_{cal}}$



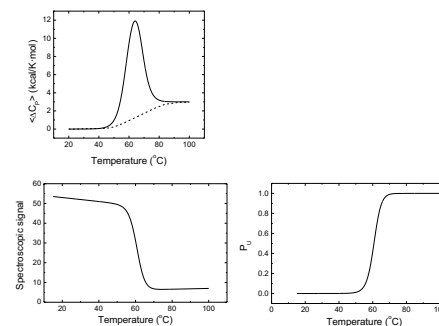
van't Hoff enthalpy

$$\Delta H_{vH} = RT^2 \frac{\partial \ln K}{\partial T}$$

$$\Delta H_{vH} = \frac{4RT_m^2 C_{P,max}}{\Delta H_{cal}}$$

$$\Delta H_{vH} = 4RT_m^2 \left(\frac{\partial \chi_U}{\partial T} \right)_{T_m}$$

model-free analysis



$$\frac{\Delta H_{vH}}{\Delta H_{cal}} = 1 \quad \text{cooperative unit} \equiv \text{molecule (single transition)}$$

$$N \leftrightarrow U$$

$$\frac{\Delta H_{vH}}{\Delta H_{cal}} < 1 \quad \text{cooperative unit} < \text{molecule (more than one transition)}$$

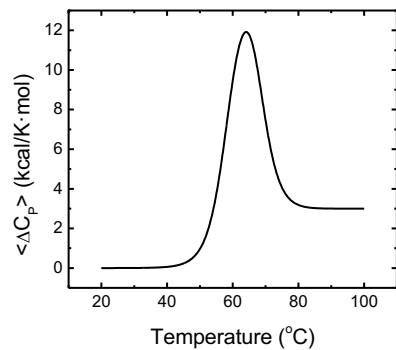
$$N \leftrightarrow I \leftrightarrow \dots \leftrightarrow U \quad \frac{\Delta H_{vH}}{\Delta H_{cal}} \neq \frac{1}{n_D}$$

$$\frac{\Delta H_{vH}}{\Delta H_{cal}} > 1 \quad \text{cooperative unit} > \text{molecule (self-associated native state)}$$

$$N_n \leftrightarrow nU \quad \frac{\Delta H_{vH}}{\Delta H_{cal}} \neq n \quad \frac{\Delta H_{vH}}{\Delta H_{cal}} = 4 \left(\frac{n - \sqrt{n}}{n - 1} \right)^2$$

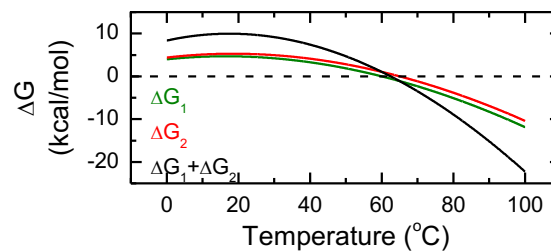
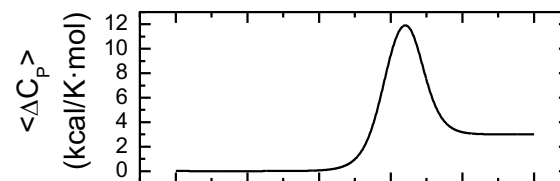
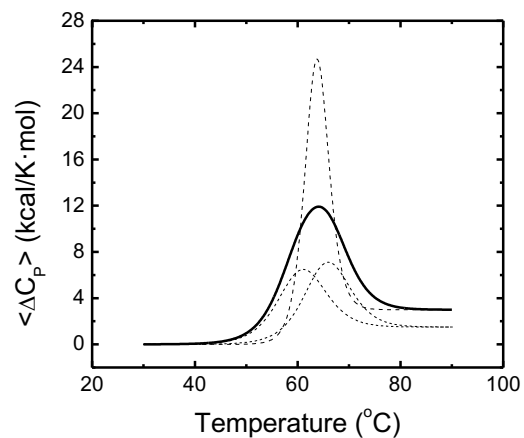
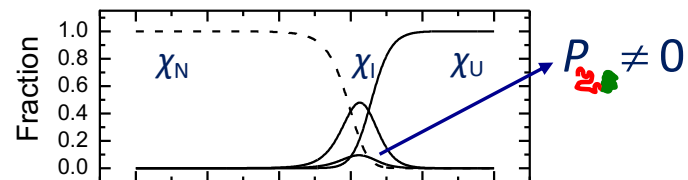


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ΔH_{cal} 144.3 kcal mol⁻¹
 T_m 63.5 °C
 $C_{p\text{max}}$ 10.2 kcal K⁻¹ mol⁻¹

ΔH_{vH} 63.4 kcal mol⁻¹
 $\Delta H_{\text{vH}}/\Delta H_{\text{cal}}$ 0.44



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Unfolding-Binding Coupling

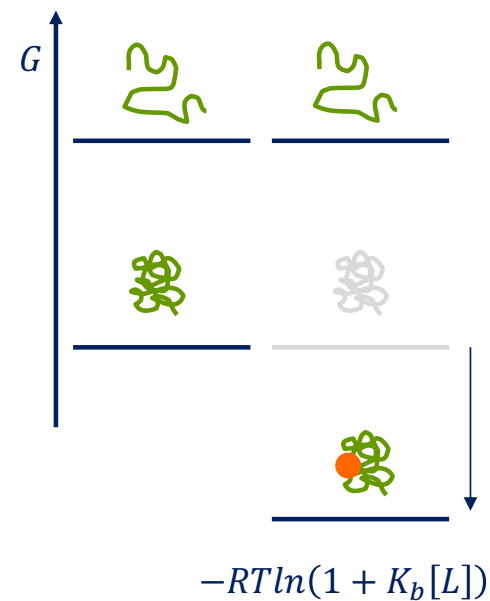
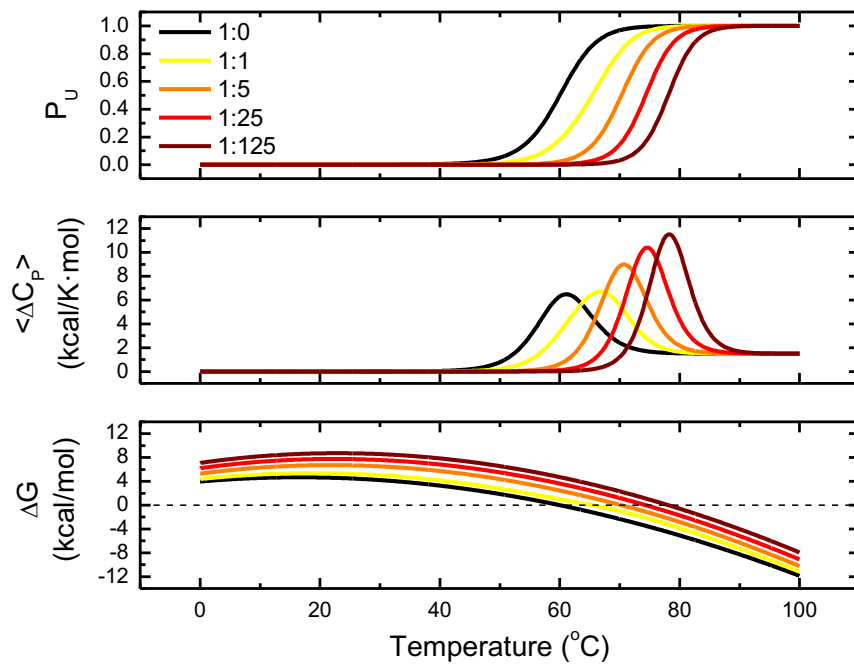


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i	ΔG_i	$\exp(-\Delta G_i/RT)$
	0	1
	ΔG_{bind}	$K_a[L]$
	ΔG^0	K^0

$$Q = 1 + K_a[L] + K^0$$





- Indirect evidence for interaction
- Determination of binding affinity (?)

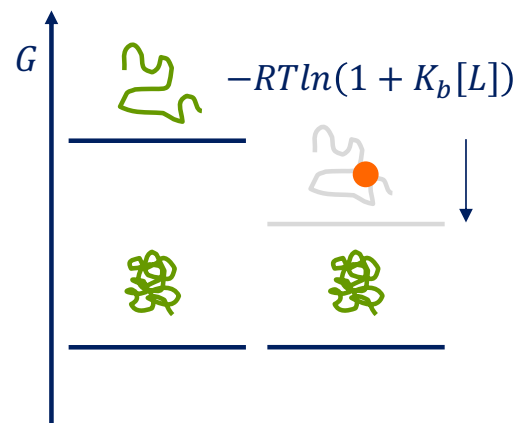
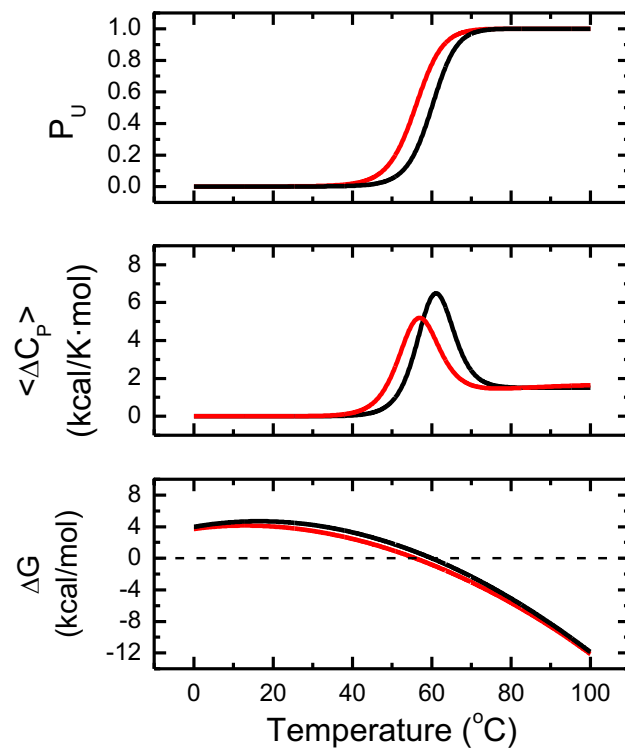


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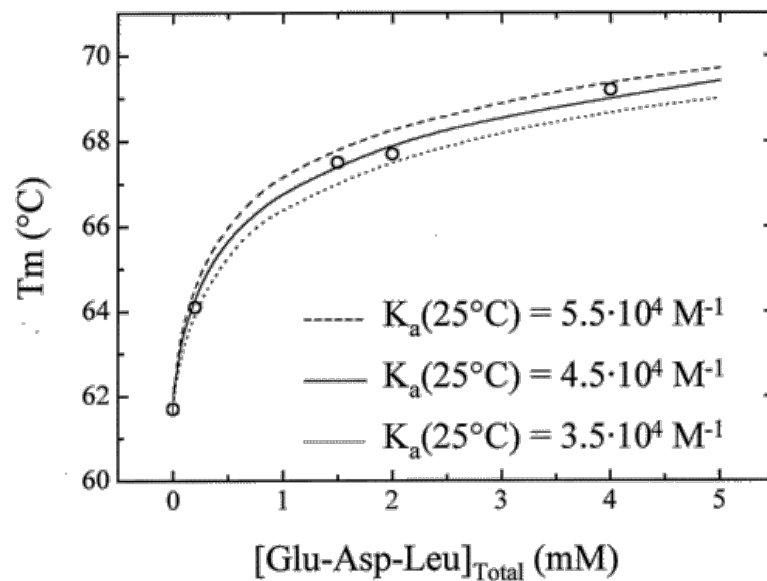
i	ΔG_i	$\exp(-\Delta G_i/RT)$
	0	1
	ΔG^0	K^0
	$\Delta G^0 + \Delta G_{\text{bind}}$	$K^0 K_a [L]$

$$Q = 1 + K^0(1 + K_a[L])$$





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$$\frac{\partial \frac{1}{T_m}}{\partial \ln[X]} \approx \frac{R}{\Delta H(T_m)} \Delta n_X$$

$$\frac{\Delta T_m}{T_m} \approx \pm \frac{n_X R T_m^0}{\Delta H(T_m)} \ln \left(1 + \frac{[X]}{K_d} \right)$$

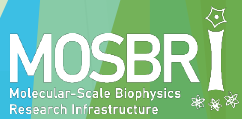


DSF

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DSF Monitoring protein unfolding using an extrinsic fluorescent reporter

Motivation:

- **Low throughput of conventional techniques (sample amount, number of assays)**
- **Conventional fluorescence plate readers do not allow broad temperature ramp**
- **Real-time qPCR allow broad temperature ramp, but do not record in the tryptophane fluorescence range**



DSF

Thermofluor® or Thermal Shift Assay (TSA)

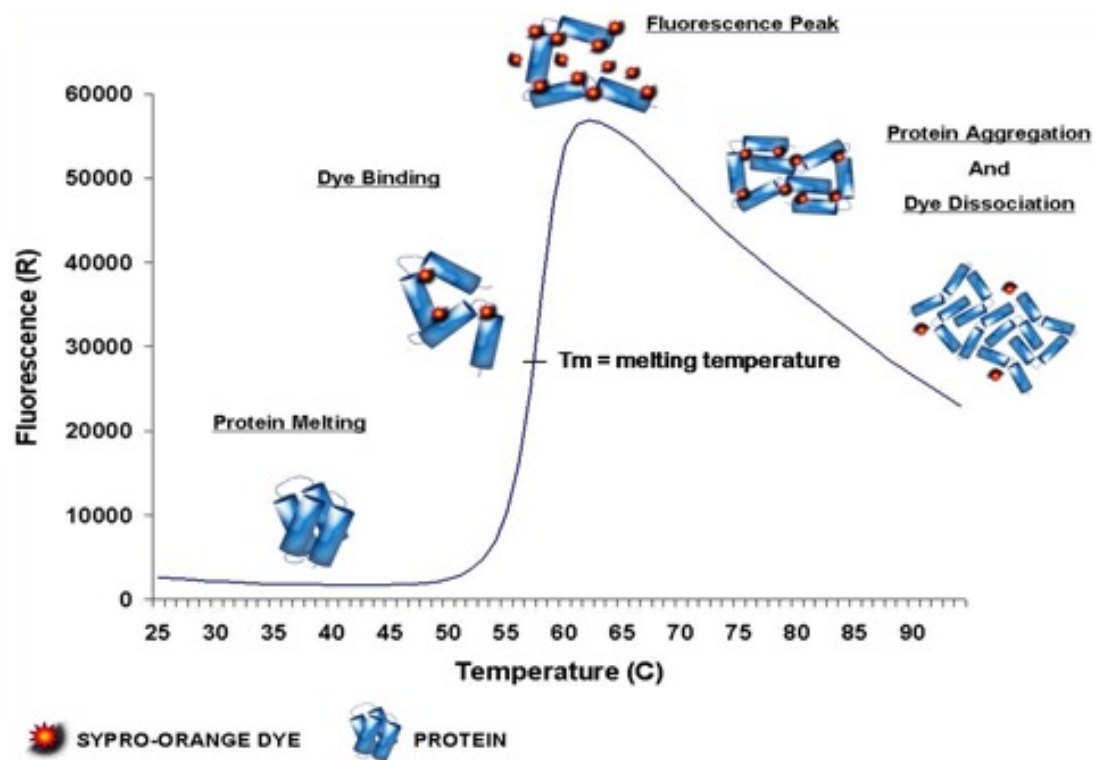
Stabilization effect induced by ligand binding or by solvent components on the thermal stability of a protein

“Differential”

Usually, differences between a “reference” sample and “test” samples are measured



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By Argonne National Laboratory -
http://www.bio.anl.gov/molecular_and_systems_biology/Sensor/sensor_images/



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DSF

Now, also using intrinsic tryptophan or cofactors fluorescence

Applications:

- Drug screening
- Drug lead optimization (**Extent of stabilization does not correlate with binding affinity!**)
- Studies of enzyme mechanism
- Protein stabilization for optimized isolation
- Characterization of engineered proteins
- Optimization of protein crystallization conditions
- Screening for inhibitors of protein-protein interactions of modulators of protein conformational changes
- Membrane proteins
- Decrypting proteins of unknown biological function
- CETSA, for cellular thermal shift assay



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Because DSF is usually employed for comparing T_m 's of samples under different conditions (e.g., with & without compound, at different pHs, with different excipients...), it is allowed to perform a minimal data analysis based on identifying the inflection point (T_{max} as a proxy for T_m).

Still, a thorough analysis can be done similar to that explained for thermal denaturations performed by conventional spectroscopy (i.e., estimation of T_m and $\Delta H(T_m)$).

